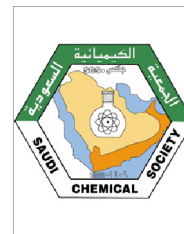




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ORIGINAL ARTICLE

Synthesis and investigation of anti-inflammatory and anticonvulsant activities of novel coumarin-diacylated hydrazide derivatives



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Abstract A number of novel coumarin derivatives synthesized by the reaction of 3-carbonyl chloride coumarin with some substituted aryl acid hydrazides to investigate their anti-inflammatory and anticonvulsant activities. Carrageenan (0.1 ml of 1%, w/v) was injected subplantarily in the right paw of rats to induce an acute model of inflammation. Anti-inflammatory efficacy was evaluated for 5 hours at 3 different dosages 5, 10, 25 mg/kg. After that, the changes in the level of paw edema volumes and percentage inhibition of all groups were observed and the most effective coumarin derivative was found as *N'*-(2-hydroxybenzoyl)-2-oxo-2H-chromene-3-carbohydrazide. In addition, *N'*-(2-oxo-2H-chromene-3-carbonyl)nicotinohydrazide, (*E*)-*N'*-(3-(4-hydroxyphenyl)acryloyl)-2-oxo-2H-chromene-3-carbohydrazide, and *N'*-(5-amino-2-hydroxybenzoyl)-2-oxo-2H-chromene-3-carbohydrazide showed their anti-inflammatory effects in a dose-dependent manner. On the other hand, pentylentetrazole (PTZ, 80 mg/kg, i.p.)-induced seizure model was used to investigate the anticonvulsant activities of six newly synthesized coumarin derivatives in mice. Hybrid compound of salicylic acid hydrazide and 3-carbonyl chloride coumarin (**8d**) was found the most promising anticonvulsant agent among all treatment groups according to the onset of seizure and survival rate. Moreover, (*E*)-*N'*-cinnamoyl-2-oxo-2H-chromene-3-carbohydrazide (**8b**) and (*E*)-*N'*-(3-(4-hydroxyphenyl)acryloyl)-2-oxo-2H-chromene-3-carbohydrazide (**8c**) has potential anticonvulsant efficiency

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in low doses (30 mg/kg). The anticonvulsant effect of these coumarin derivatives may be through enhanced GABA-mediated inhibition in the brain.

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1. Introduction

Epilepsy is a serious chronic neurologic disorder which characterized by spontaneously occurring seizures. It can be seen among people of all ages. Currently available antiepileptic drugs (AEDs) are able to successfully control seizures with in most patients with epilepsy, however, 30–40% of all epileptic patients still suffer from some side effects and seizure resistance to the current AEDs (Sørensen and Kokaia, 2013). Therefore, researchers keep to continue to develop novel anti-convulsant compounds to cover seizures to treat epilepsy.

Inflammation is the natural defense mechanism of the body against irritants, trauma, pathogens, or microbial invasion. It occurs in the human body with symptoms such as pain, swelling, redness, heat, and disturbance of function (Aghasafari et al., 2019; Minhas et al., 2017; Pahwa and Jialal, 2019). Migration and activation of leukocytes cause tissue destruction and enhanced blood vessel permeability. As a result, inflammation could be occur by activation of several enzymes (lipoxygenases and cyclooxygenases (COX)), production of inflammatory mediators (leukotrienes and prostaglandins), and reactive oxygen species (Abdel-Lateff et al., 2020).

The development of hybrid molecules via the combination of two or more different pharmacophore units in one frame is an attractive area in the design of new drugs. Hence, these new molecules may show interesting biological activities. Due to multifunctional features of coumarin among the new hybrid molecules, extensive efforts have been made on design and synthesis of coumarin derivatives, which was coupled with different bioactive molecules (Sashidhara et al., 2011; Sashidhara et al., 2010).

Coumarins are widely reported that they have several biological activities. They have been used to treat diseases in a wide range from cancer, burns, brucellosis to cardiovascular and rheumatic diseases (Kennedy and Thornes, 1997; Hoult

and Payá, 1996; Ma et al., 2008; Kostova, 2005; Musa and Cooperwood, 2008). In addition, various coumarin derivatives are reported that they have anti-inflammatory and antiepileptic activities. They also able to inhibit the lipoxygenase and cyclooxygenase enzymes (Piller, 1975). Coumarin compounds, especially substituted with hydroxyl or other electron releasing groups were found to inhibit lipid peroxidation and to scavenge hydroxyl radicals and superoxide anions (Payá et al., 1992). α -lipoic acid derivatives of coumarin-3-carboxamides were also reported as potent antioxidant and anti-inflammatory agents (Kontogiorgis and Hadjipavlou-Litina, 2005; Melagraki et al., 2009; Sashidhara et al., 2011; Vardhan Reddy et al., 2016).

Diacylhydrazines have been reported as a powerful non-teroidal insect growth regulators and the first commercialized product RH-5992 (tebufenozide) was reported by Rohm&Haas Company (Heller et al., 1992). Later, replacement of one of aryl units with a benzoheterocycle containing oxygen units gave the products ANS-118 and JS-118 (Li et al., 2010; Zhang et al., 2001). CUI et al synthesized a series of novel diacylhydrazine derivatives containing a furan ring and investigated their anti-tumor activity. They reported that some of the furan-*N,N'*-diacylhydrazites have potential activity against some human cancer cell lines, such as human leukemia cell line HL-60, human gastric carcinoma cell line (Cui et al., 2008).

After all mentioned findings, novel diacyl hydrazide derivatives containing biaryl unites which one of them is coumarin have been designed to evaluate their anti-inflammatory and anticonvulsant activities. During the design of the molecules, keeping in mind the synergistic effects of two biologically active units in one frame, we considered two hydrogen bonding and two electron donor group ($-\text{NHCO}-$) and two hydrophobic aryl groups (coumarin and aryl acid) as key pharmacophores for anticonvulsant and anti-inflammatory activities (Fig. 1). Six novel *N,N'*-diacyl hydrazide derivatives (**8a**-

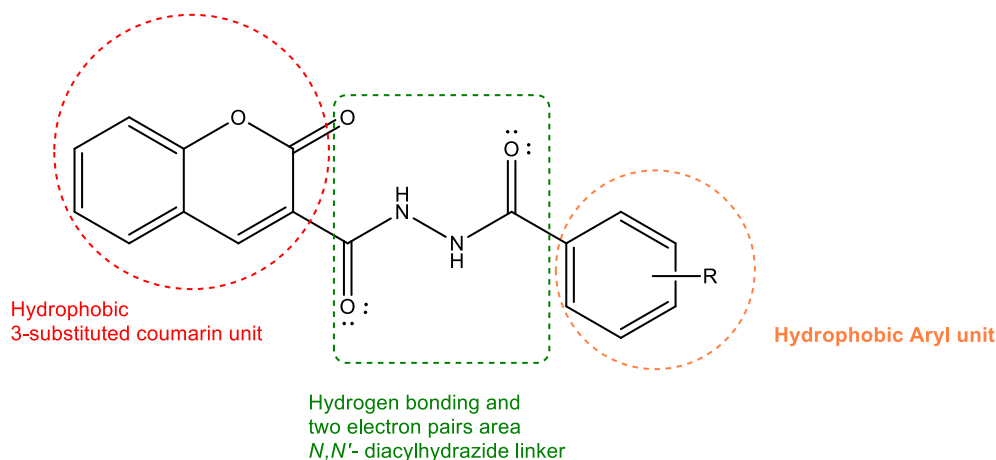
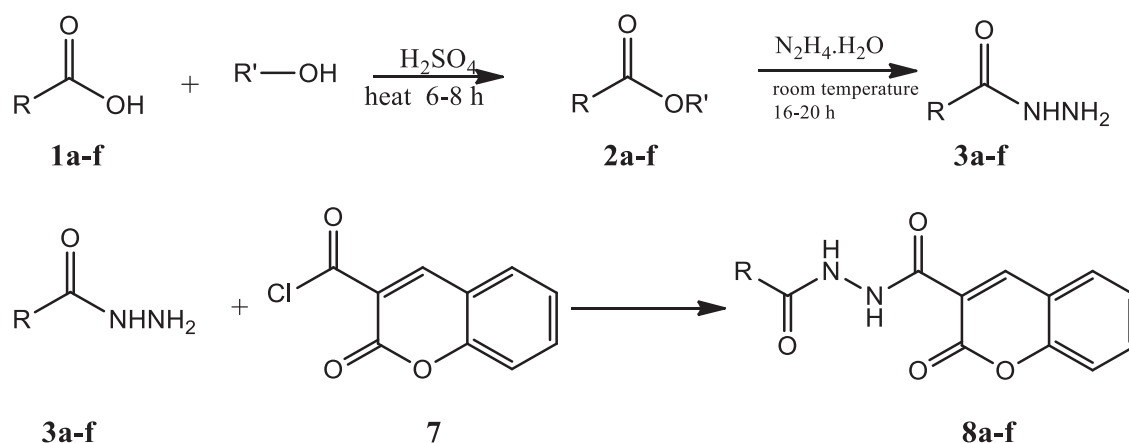


Fig. 1 Key pharmacophore units of novel coumarin-diacylhydrazide hybrids.



Scheme 1 Synthesis of diacylated hydrazides hybrids.

f) were prepared by the reaction of 2-oxo-2H-chromen-3-carbonyl chloride (**7**) with some substituted aryl acid hydrazides (**3a-f**) (Scheme 1).

2. Materials and methods

2.1. Chemistry

All the reagents were obtained from different commercial sources. Unless noted otherwise, all of compounds were used as provided without further purification. Nicotinic acid ($\geq 98\%$), salicylaldehyde ($\geq 98\%$), *trans*-cinnamic acid ($\geq 99\%$), *p*-coumaric acid ($\geq 98\%$), salicylic acid (ACS reagent $\geq 99\%$), 5-amino-salicylic acid (95–98%), gallic acid (97%), hydrazine hydrate (reagent grade, 50–60%), malonic acid (99%), thionyl chloride (reagent grade, 97%), sulphuric acid (99.999%), acetic acid glacial (reagent grade $\geq 99\%$), tetrahydrofuran ($\geq 99\%$), methanol (anhydrous, 99.8%) and ethanol (99.8%) were purchased from Sigma Aldrich. λ -carrageenan, pentylene tetrazole (PTZ), indomethacin, and carbamazepine were purchased from Sigma-Aldrich (Poole, UK). Follow up of the reactions and checking the purity of the compounds were made by TLC on silica gel-precoated aluminium sheets (Type 60, F₂₅₄, Merck, Darmstadt, Germany) using hexane/ethyl acetate 80–20 (4:1, v/v) and the spots were detected by exposure to UV lamp at λ_{254} nanometer for few seconds. Melting points were determined using an Electrothermal 9100 instrument in open capillary tubes. IR spectra were recorded on a Perkin-Elmer 55,148 spectrometer using KBr pellets. ¹H and ¹³C NMR spectra were recorded in CDCl₃ or DMSO-*d*₆ [using the solvent peak as internal reference (DMSO-*d*₆: δ H 2.50; δ C 39.51 and CDCl₃ at 7.27 ppm for ¹H and 77.0 ppm for ¹³C) on a Bruker Avance III 400 MHz spectrometer operating at 400 MHz and 100 MHz, respectively. All chemical shift values are quoted in ppm and coupling constants quoted in Hz. Multiplicities are indicated, s (singlet), d (doublet), t (triplet), q (quartet), sept (septet), m (multiplet), br s (broad singlet). Elemental analyses were measured on a Thermo Flash 2000 Organic Elemental Analyzer. The structural data for 2a [Erharuyi et al. \(2015\)](#); 2b [Steele et al. \(2020\)](#); 2c [Lee et al. \(2013\)](#); 2d [Petrus and Sobota, 2013](#); 2e [Fiuza et al. \(2004\)](#); 2f [Bailey et al. \(2004\)](#); 3a [Vimal Kumar Varma and Amareshwar \(2011\)](#); 3b [Katritzky et al. \(2001\)](#);

3c [Saritha and Rajitha \(2019\)](#); 3d [Pattan et al. \(2009\)](#); 3e [Taha et al. \(2019\)](#); 3f [Ersan et al. \(2008\)](#); 6 [Patil et al. \(2018\)](#); 7 [Zobaydi et al. \(2016\)](#) and 8d [Jesumoroti et al. \(2019\)](#) is in accordance with the results reported in the literature.

2.1.1. Synthesis of carboxylic acid ester derivatives (2a–2f) ([Fiuza et al., 2004](#))

For the esterification reaction, the reaction was carried out with carboxylic acid and methanol taken at a ratio of 1:10 under reflux for about 6 h at about 60–80 °C. After completion of the reaction, excess of methyl alcohol was distilled and the solid formed was washed with cold water and purified from impurities. The solid was dissolved in ethanol, water was added until the cloudy color was achieved and allowed to crystallize. The product was identified using FT-IR and NMR spectroscopy.

2.1.1.1. Methyl nicotinate (2a) ([Erharuyi et al., 2015](#)). white solid; yield 1.165 g (85%); mp (°C): 42–44; FT-IR (KBr) 3030 (CH, aromatic), 2967 (CH, aliphatic), 1710 (C=O), 1674 (C=O) 1606, 1567 (C=C, aromatic), 1281 (C–O); δ H (400 MHz, DMSO-*d*₆) 9.79 (s, 1H), 7.05 (d, *J* = 8 Hz, 1H), 6.87–6.83 (m, 1H), 6.72 (d, *J* = 8 Hz, 1H), 3.86 (s, 3H); δ C (100 MHz, DMSO-*d*₆) 169.68, 151.81, 140.66, 123.30, 117.61, 113.12, 52.22; Anal. Calcd. for C₇H₇NO₂: C, 61.31; H, 5.14; N, 10.21 Found: C, 61.50; H, 5.11; N, 10.17.

2.1.1.2. Methyl cinnamate (2b) ([Steele et al., 2020](#)). Pale yellow solid; yield 1.525 g (94%); mp (°C): 125–127; FT-IR (KBr) 3111 (CH, aromatic), 2952 (CH, aliphatic), 1682 (C=O), 1632 (C=C, aliphatic), 1597, 1514 (C=C, aromatic), 1193 (C–O), 984 (C=C, aliphatic-trans), 831 (C–H, *p*-substitue); δ H (400 MHz, DMSO-*d*₆) 7.38 (d, *J* = 16 Hz, 1H), 7.21 (d, *J* = 16 Hz, 1H), 7.10–6.96 (m, 2H), 6.74–6.64 (m, 3H), 3.90 (s, 3H); δ C (100 MHz, DMSO-*d*₆) 163.97, 156.66, 134.15, 132.23, 131.18, 127.87, 116.58, 63.75; Anal. Calcd. for C₁₀H₁₀O₂: C, 74.06; H, 6.21; Found: C, 74.21; H, 6.17.

2.1.1.3. (E)-methyl 3-(4-hydroxyphenyl)acrylate (2c) ([Lee et al., 2013](#)). Yellow solid; yield 1.337 g (65%); mp (°C): 143–145; FT-IR (KBr) 3356 (O–H), 3039 (CH, aromatic), 2952

(CH, aliphatic), 1682 (C=O) 1632 (C=C, aliphatic), 1597, 1514 (C=C, aromatic), 1328 (C—O), 984 (C=C, aliphatic-trans), 830 (C—H, p-substitue); δ H (400 MHz, DMSO- d_6) 8.96 (s, 1H), 7.21 (d, J = 16 Hz, 1H), 7.08 (d, J = 16 Hz, 1H), 6.97 (d, J = 8 Hz, 2H) 6.65 (d, J = 8 Hz, 2H), 3.85 (s, 3H); δ C (100 MHz, DMSO- d_6) 170.93, 155.44, 145.54, 131.19, 128.99, 115.01, 114.88, 61.56; Anal. Calcd. for $C_{10}H_{10}O_3$: C, 67.41; H, 5.66; Found: C, 67.57; H, 5.62.

2.1.1.4. Methyl 2-hydroxybenzoate (2d) (Petrus and Sobota, 2013). clear liquid; yield 1.370 g (90%); FT-IR (KBr) 3152 (CH, aromatic), 2955 (CH, aliphatic), 1674 (C=O), 1614, 1487 (C=C, aromatic), 1302 (C—O), 754, 699 (C—H, o-substitue); δ H (400 MHz, DMSO- d_6) 10.55 (s, 1H), 7.78 (d, J = 8 Hz, 1H), 7.53 (t, J = 8 Hz, 1H), 6.92–6.89 (m, 2H), 3.88 (s, 3H); δ C (100 MHz, DMSO- d_6) 169.31, 160.14, 135.59, 129.87, 119.29, 117.31, 112.76, 52.33; Anal. Calcd. for $C_8H_8O_3$: C, 63.15; H, 5.30; Found: C, 63.29; H, 5.27.

2.1.1.5. Methyl 3,4,5-trihydroxybenzoate (2e) (Fiuza et al., 2004). white solid; yield 1.842 g (99.99%); mp (°C) : 201–203; FT-IR (KBr) 3355 (O—H), 3045 (C—H, aromatic), 2952 (C—H, aliphatic), 1674 (C=O), 1614, 1585 (C=C, aromatic), 1300 (C—O); δ H (400 MHz, DMSO- d_6) 9.30 (s, 2H), 8.31 (s, 1H), 6.79 (s, 2H), 3.74 (s, 3H); δ C (100 MHz, DMSO- d_6) 166.45, 145.40, 136.40, 123.17, 106.23, 54.45; Anal. Calcd. for $C_8H_8O_5$: C, 52.18; H, 4.38; Found: C, 52.39; H, 4.35.

2.1.1.6. Methyl 5-amino-2-hydroxybenzoate (2f) (Bailey et al., 2004). white solid; yield 1.504 g (90%); mp (°C): 95–97; FT-IR (KBr) 3409 (O—H), 3326 (N—H), 3027 (C—H, aromatic), 2962 (C—H, aliphatic), 1670 (C=O), 1613, 1594 (C=C, aromatic), 1340 (C—N), 1207 (C—O), 890 (C—H, 1,2,4-tri substitue); δ H (400 MHz, DMSO- d_6) 11.13 (s, 2H), 9.01 (s, 1H), 7.70 (d, J = 8 Hz, 1H), 7.41 (t, J = 8 Hz, 1H), 6.99 (d, J = 8 Hz, 1H), 3.34 (s, 3H); δ C (100 MHz, DMSO- d_6) 168.53, 157.94, 134.45, 128.03, 120.21, 117.90, 113.53, 54.39; Anal. Calcd for $C_8H_9NO_3$: C, 57.48; H, 5.43; N, 8.38; Found: C, 57.64; H, 5.40; N, 8.33.

2.1.2. Synthesis of carboxylic acid hydrazide derivatives (3a-3f) (Ersan et al., 2008)

The ester compounds (5 mmol) synthesized in the first step were dissolved in methanol (50 mmol, 2.02 ml) and an equivalent amount of hydrazine hydrate was added thereto. The reaction was carried out by mixing the mixture at room conditions for 16–20 h. The solid obtained as a result of the reaction was removed from the impurities by washing with methanol in the filtration apparatus. The crude product was recrystallized from ethanol to obtain pure product. The product was identified using FT-IR and NMR spectroscopy.

2.1.2.1. Nicotinohydrazide (3a) (Vimal Kumar Varma and Amareshwar, 2011). white solid; yield 0.460 g (67%); mp (°C): 200–202; FT-IR (KBr) 3367 (N—H), 3279 (NH₂), 3029 (C—H, aromatic), 2909, 2867 (C—H, aliphatic), 1710 (C=O), 1612, 1566 (C=C, aromatic), 1226 (C—N), 756 (C—H, m-substitue); δ H (400 MHz, DMSO- d_6) 9.36 (s, 1H), 7.98 (s, 1H), 7.05 (d, J = 8 Hz, 1H), 6.87 (d, J = 8 Hz, 1H), 6.84–6.71 (m, 1H), 4.33 (s, 2H); δ C (100 MHz, DMSO- d_6) 168.48,

150.23, 138.95, 121.46, 116.43, 112.13; Anal. Calcd. for $C_6H_7N_3O$: C, 52.55; H, 5.14; N, 30.64; Found: C, 52.76; H, 5.10; N, 30.58.

2.1.2.2. Cinnamohydrazide (3b) (Katritzky et al., 2001). Brown solid; yield 0.617 g (76%); mp (°C): 114–116; FT-IR (KBr) 3267 (NH₂), 3187 (N—H), 3011 (C—H, aromatic), 2973 (C—H, aliphatic), 1650 (C=O), 1636 (C=C, aliphatic), 1597, 1514 (C=C, aromatic), 1279 (C—N), 1248 (C—O) 965 (C=C, aliphatic-trans), 826 (C—H, p-substitue); δ H (400 MHz, DMSO- d_6) 8.30 (s, 1H), 7.72 (t, J = 8 Hz, 4H), 7.42 (d, J = 16 Hz, 1H), 7.30 (d, J = 8 Hz, 1H), 6.82 (d, J = 16 Hz, 1H), 3.83 (s, 2H); δ C (100 MHz, DMSO- d_6) 163.89, 154.36, 140.89, 132.95, 128.46, 118.08, 115.85; Anal. Calcd. for $C_9H_{10}N_2O$: C, 66.65; H, 6.21; N, 17.27; Found: C, 66.84; H, 6.17; N, 17.22.

2.1.2.3. (E)-3-(4-hydroxyphenyl)acrylohydrazide (3c) (Saritha and Rajitha, 2019). white solid; yield 0.642 g (72%); mp (°C): 143–145; FT-IR (KBr) 3267 (O—H), 3187 (N—H), 3011 (C—H, aromatic), 2943 (C—H, aliphatic), 1650 (C=O), 1637 (C=C, aliphatic), 1585, 1515 (C=C, aromatic), 1279 (C—N), 1248 (C—O) 965 (C=C, aliphatic-trans), 825 (C—H, p-substitue); δ H (400 MHz, DMSO- d_6) 8.96 (s, 1H), 8.17 (s, 1H), 7.20 (d, J = 16 Hz, 1H), 7.09 (d, J = 16 Hz, 1H), 6.97 (d, J = 8 Hz, 2H), 6.68 (d, J = 8 Hz, 2H), 4.46 (s, 2H); δ C (100 MHz, DMSO- d_6) 168.67, 153.43, 147.86, 130.36, 128.73, 118.29, 116.08; Anal. Calcd. for $C_9H_{10}N_2O_2$: C, 60.66; H, 5.66; N, 15.72; Found: C, 60.85; H, 5.63; N, 15.66.

2.1.2.4. 2-hydroxybenzohydrazide (3d) (Pattan et al., 2009). white solid; yield 0.555 g (73%); mp (°C): 149–151; FT-IR (KBr) 3340 (—OH), 3190 (N—H), 1656 (C=O), 1531, 1457 (C=C, aromatic), 1332 (C—N), 1291 (C—O), 769, 698 (C—H, o-substitue); δ H (400 MHz, DMSO- d_6) 8.96 (s, 1H), 8.17 (s, 1H), 7.22–6.95 (m, 2H), 6.80–6.64 (m, 2H), 4.44 (s, 2H); δ C (100 MHz, DMSO- d_6) 163.97, 155.43, 134.08, 131.18, 128.98, 127.87, 114.92; Anal. Calcd. for $C_7H_8N_2O_2$: C, 55.26; H, 5.30; N, 18.41; Found: C, 55.45; H, 5.27; N, 18.36.

2.1.2.5. 3,4,5-trihydroxybenzohydrazide (3e) (Taha et al., 2019). white solid; yield 0.875 g (95%); mp (°C): 283–285; FT-IR (KBr) 3424 (O—H), 3390 (N—H), 3297 (NH₂), 3143 (C—H, aromatic), 1656 (C=O), 1602, 1539 (C=C, aromatic), 1338 (C—N), 1204 (C—O); δ H (400 MHz, DMSO- d_6) 9.33 (s, 1H), 8.38 (s, 3H), 6.75 (s, 2H), 4.41 (s, 2H); δ C (100 MHz, DMSO- d_6) 166.46, 145.48, 136.40, 123.30, 106.40; Anal. Calcd. for $C_7H_8N_2O_4$: C, 45.66; H, 4.38; N, 15.21; Found: C, 45.83; H, 4.34; N, 15.15.

2.1.2.6. 5-amino-2-hydroxybenzohydrazide (3f) (Ersan et al., 2008). white solid; yield 0.610 g (73%); mp (°C): 190–192; FT-IR (KBr) 3337 (O—H), 3196 (N—H), 3027 (C—H, aromatic), 1585 (C=O), 1539, 1470 (C=C, aromatic), 1315 (C—N), 1217 (C—O), 900 (C—H, 1,2,4-tri substitue); δ H (400 MHz, DMSO- d_6) 11.41 (s, 2H), 10.23 (s, 1H), 9.32 (s, 1H), 7.04–6.73 (m, 3H), 4.71 (s, 2H); δ C (100 MHz, DMSO- d_6) 170.92, 155.43, 145.53, 134.17, 131.19, 128.99, 115.03; Anal. Calcd. for $C_7H_9N_3O_2$: C, 50.29; H, 5.43; N, 25.14; Found: C, 50.49; H, 5.40; N, 25.09.

2.1.3. Synthesis of 2-oxo-2H-chromene-3-carboxylic acid (6) (Karade et al., 2007)

Malonic acid (10 mmol) (4), salicylaldehyde (10 mmol) (5) and L-proline (10% by weight) were added to the 50 ml round-bottom flask and the reaction was carried out by mixing with a magnetic stirrer at 80–90 °C for 30 min. The solid formed was filtered and washed away with purified water to remove impurities. The crude product was recrystallized from ethanol to obtain pure product. The product was identified using FT-IR and NMR spectroscopy.

2.1.3.1. 2-oxo-2H-chromene-3-carboxylic acid (6) (Patil et al., 2018). white solid; yield 1.711 g (90%); mp (°C): 185–187; FT-IR (KBr) 3057 (O—H), 3005 (C—H, aromatic), 2912 (C—H, aliphatic), 1737 (C=O), 1606, 1557 (C=C, aromatic), 1226 (C—O); δ H (400 MHz, DMSO- d_6) 13.69 (s, 1H), 8.71 (s, 1H), 7.87 (d, J = 8 Hz, 2H), 7.69 (t, J = 8 Hz, 1H), 7.36 (t, J = 8 Hz, 1H); δ C (100 MHz, DMSO- d_6) 163.74, 156.68, 154.37, 148.32, 134.19, 130.10, 124.72, 118.12, 117.87, 116.01; Anal. Calcd. for $C_{10}H_6O_4$: C, 63.16; H, 3.18; Found: C, 63.37; H, 3.15.

2.1.4. Synthesis of 2-oxo-2H-chromene-3-carbonyl chloride (7) (Zobaydi et al., 2016)

3-carboxy coumarin (10 mmol) and thionyl chloride (20 mmol) were added to the 50 ml round bottom flask and mixed under reflux for 6–8 h in solvent-free medium. The solid formed was filtered and washed away with purified water to remove impurities. The crude product was recrystallized from ethanol to obtain pure product. The product was identified using FT-IR and NMR spectroscopy.

2.1.4.1. 2-oxo-2H-chromene-3-carbonyl chloride (7) (Zobaydi et al., 2016). yellow solid; yield 1.817 g (95%); mp (°C): 155–157; FT-IR (KBr) 3058 (C—H, aromatic), 1769 (C=O), 1605, 1556 (C=C, aromatic), 1364 (C—Cl), 1287 (C—O), 758 (C—Cl); δ H (400 MHz, DMSO- d_6) 8.71 (s, 1H), 7.89 (d, J = 8 Hz, 1H), 7.85 (d, J = 8 Hz, 1H), 7.72–7.34 (m, 2H); δ C (100 MHz, DMSO- d_6) 160.30, 159.33, 153.11, 147.90, 136.19, 128.13, 125.03, 120.35, 118.61, 115.22; Anal. Calcd. for $C_{10}H_5ClO_3$: C, 57.58; H, 2.42; Cl, 17.00 Found: C, 57.79; H, 2.39; Cl, 16.54.

2.1.5. Synthesis of coumarin hydrazide hybrid derivatives (8a–8f)

Carboxylic acid hydrazide derivative (10 mmol) was taken into a 50 ml round bottom flask and 3–4 drops of acetic acid was added by dissolving with THF and mixed. 2-oxo-2H-chromene-3-carbonyl chloride (3b) (10 mmol) was added on it and mixed under reflux for 6 h. After the reaction was completed, the excess of THF was distilled and removed. The remaining solid was neutralized with aqueous Na_2CO_3 . The crude product was recrystallized from ethanol to obtain pure product. The product was identified using FT-IR and NMR spectroscopy.

2.1.5.1. N' -(2-oxo-2H-chromene-3-carbonyl)nicotinohydrazide (8a). white solid; yield 2.784 g (90%); mp (°C): >300; FT-IR (KBr) 3365, 3282 (N—H), 3060 (C—H, aromatic), 2923, 2853 (C—H, aliphatic), 1737, 1710, 1643 (C=O), 1612, 1566 (C=C, aromatic), 1314 (C—N), 1226 (C—O), 766 (C—H, m-

substitute), 756 (C—H o-substitute); δ H (400 MHz, DMSO- d_6) 9.79 (s, 1H), 9.14 (s, 1H), 8.73 (s, 1H), 8.41–8.22 (m, 3H), 6.87–6.71 (m, 5H); δ C (100 MHz, DMSO- d_6) 163.93, 156.65, 154.41, 148.33, 134.22, 130.13, 127.69, 124.75, 118.25, 117.92, 116.06, 112.84; Anal. Calcd. for $C_{16}H_{11}N_3O_4$: C, 62.14; H, 3.58; N, 13.59; Found: C, 62.35; H, 3.55; N, 13.53.

2.1.5.2. (*E*)- N' -cinnamoyl-2-oxo-2H-chromene-3-carbohydrazide (8b). yellow solid; yield 2.842 g (85%); mp (°C): >300; FT-IR (KBr) 3367 (N—H), 3267 (NH₂), 3061 (C—H, aromatic), 2952, 2838 (C—H, aliphatic), 1710 (C=O), 1637 (C=C, aliphatic), 1611, 1514 (C=C, aromatic), 1248 (C—N), 1171 (C—O) 965 (C=C, aliphatic-trans), 826 (C—H, p-substitute); δ H (400 MHz, DMSO- d_6) 9.01 (s, 1H), 8.60 (s, 1H), 7.88 (d, J = 16 Hz, 1H), 7.74–7.69 (m, 5H), 7.43–7.36 (m, 5H), 7.14 (d, J = 16 Hz, 1H); δ C (100 MHz, DMSO- d_6) 165.87, 158.47, 152.50, 140.21, 136.62, 132.58, 126.32, 119.90, 117.04, 115.28; Anal. Calcd. for $C_{19}H_{14}N_2O_4$: C, 68.26; H, 4.22; N, 8.38; Found: C, 68.48; H, 4.19; N, 8.32.

2.1.5.3. (*E*)- N' -(3-(4-hydroxyphenyl)acryloyl)-2-oxo-2H-chromene-3-carbohydrazide (8c). brown solid; yield 2.452 g (70%); mp (°C): >300; FT-IR (KBr) 3634 (O—H), 3324 (N—H), 3200 (NH₂), 3028 (C—H, aromatic), 2955, 2897 (C—H, aliphatic), 1653 (C=O), 1620 (C=C, aliphatic), 1603, 1508 (C=C, aromatic), 1260 (C—N), 1242 (C—O) 997 (C=C, aliphatic-trans), 825 (C—H, p-substitute); δ H (400 MHz, DMSO- d_6) 13.27 (s, 1H), 8.73 (s, 2H), 8.45 (s, 1H), 7.88 (d, J = 16 Hz, 1H), 7.74–7.69 (m, 2H), 7.43–7.36 (m, 4H), 7.13 (d, J = 16 Hz, 1H); δ C (100 MHz, DMSO- d_6) 163.93, 156.64, 154.421, 148.33, 134.21, 130.13, 124.75, 118.25, 117.92, 116.06; Anal. Calcd. for $C_{19}H_{14}N_2O_5$: C, 65.14; H, 4.03; N, 8.00; Found: C, 65.35; H, 4.00; N, 7.94.

2.1.5.4. N' -(2-hydroxybenzoyl)-2-oxo-2H-chromene-3-carbohydrazide (8d) (Jesumoroti et al., 2019). yellow solid; yield 2.919 g (90%); mp (°C): >300; FT-IR (KBr) 3361 (OH), 3265 (N—H), 3053 (C—H, aromatic), 1698 (C=O), 1609, 1564 (C=C, aromatic), 1287 (C—N), 1221 (C—O), 769, 698 (C—H, o-substitute); δ H (400 MHz, DMSO- d_6) 11.14 (s, 1H), 9.01 (s, 1H), 7.70 (d, J = 8 Hz, 2H), 7.44–7.38 (m, 2H), 6.99 (d, J = 8 Hz, 4H); δ C (100 MHz, DMSO- d_6) 162.75, 158.61, 154.23, 141.56, 133.24, 130.80, 128.38, 124.75, 119.59, 118.17, 116.51; Anal. Calcd. for $C_{17}H_{12}N_2O_5$: C, 62.96; H, 3.73; N, 8.64; Found: C, 63.20; H, 3.69; N, 8.57.

2.1.5.5. 2-oxo- N' -(3,4,5-trihydroxybenzoyl)-2H-chromene-3-carbohydrazide (8e). yellow solid; yield 3.385 g (95%); mp (°C): >300; FT-IR (KBr) 3536 (O—H), 3115 (N—H), 3053 (C—H, aromatic), 2930 (C—H, aliphatic), 1687 (C=O), 1605, 1556 (C=C, aromatic), 1348 (C—N), 1207 (C—O); δ H (400 MHz, DMSO- d_6) 10.57 (s, 1H), 10.44 (s, 1H), 9.19 (s, 2H), 8.91 (s, 1H), 8.83 (s, 1H), 8.03 (d, J = 8 Hz, 1H), 7.78 (t, J = 8 Hz, 1H), 7.55–7.45 (m, 2H); δ C (100 MHz, DMSO- d_6) 165.04, 159.82, 159.93, 153.94, 147.98, 145.48, 137.05, 134.36, 130.33, 125.19, 122.07, 116.21, 118.40, 118.27, 107.11; Anal. Calcd. for $C_{17}H_{12}N_2O_7$: C, 57.31; H, 3.39; N, 7.86; Found: C, 57.50; H, 3.36; N, 7.80.

2.1.5.6. N' -(5-amino-2-hydroxybenzoyl)-2-oxo-2H-chromene-3-carbohydrazide (8f). brown solid; yield 2.714 g (80%); mp (°C): >300; FT-IR (KBr) 3382 (O—H), 3265 (N—H), 3052

(C—H, aromatic), 1698 (C=O), 1608, 1583 (C=C, aromatic), 1287 (C—N), 1220 (C—O), 922 (C—H, 1,2,4-tri substitue); δ H (400 MHz, DMSO- d_6) 10.88 (s, 1H), 9.08 (s, 1H), 8.92 (s, 1H), 8.75 (s, 1H), 8.03 (d, J = 8 Hz, 1H), 7.77 (d, J = 8 Hz, 1H), 7.58–7.45 (m, 4H); δ C (100 MHz, DMSO- d_6) 163.33, 159.71, 159.69, 153.95, 152.26, 148.54, 147.92, 135.26, 134.35, 130.33, 128.32, 125.19, 123.55, 118.45, 118.29, 116.22; Anal. Calcd. for $C_{17}H_{13}N_3O_5$: C, 60.18; H, 3.86; N, 12.38; Found: C, 60.39; H, 3.82; N, 12.32.

2.2. *In vivo* animal test

Experimental protocol of *in vivo* animal tests was approved by Marmara University Animal Experiments Local Ethics Committee (permission number: 57.2019.mar). Adult Sprague-Dawley rats of either sex at the age of 3–4 month and weighing 300–450 g for anti-inflammatory activity test, adult BALB/c mice of either sex at the age of 2–3 month and weighing 20–30 g for anticonvulsant activity were obtained from The Experimental Animal Implementation and Research Center (DEHAMER) of Marmara University. The rats and mice were maintained in controlled temperature (20 ± 2 °C), humidity (40–60%), and 12 h dark/light cycle-regulated rooms, with free access to food and top water. All experiments were performed at the fixed hours between 09:00–19:00 during the light phase of the day. All necessary precautions were taken before the experiment and the factors that would adversely affect the parameters were minimized during the study. The experimental groups were chosen randomly after the adaptation period of 2 days for mice.

2.2.1. Experimental design

2.2.1.1. Experimental design for anti-inflammatory activity. Anti-inflammatory and anticonvulsant activities for six different coumarin derivatives were evaluated and compared with *in vivo* animal test. The experimental rats for anti-inflammatory activity test were divided into 20 groups each consisting of 6 rats of either sex (equal ratio). All synthesized coumarin derivatives were suspended in 0.5% methylcellulose (vehicle for all coumarin derivatives) and given using oral gavage for once (5 ml/kg). Group A is model control group was given vehicle; Group B is positive control group was given indomethacin (10 mg/kg (Cong et al., 2015) suspended in 0.5% methylcellulose. All coumarin derivatives were given to rats in three different dosages (5, 10, and 25 mg/kg (Witaicenis et al., 2014) (Hemshekhkar et al., 2013) for evaluating their anti-inflammatory activities. Group **8a1-8a3** were given treatment 1 in low, middle, and high doses, respectively. In the same way, group **8b1-8b3** were given treatment 2, group **8c1-8c3** were given treatment 3, group **8d1-8d3** were given treatment 4, group **8e1-8e3** were given treatment 5, and group **8f1-8f3** were given treatment 6 in three different doses from low to high, respectively.

2.2.2. Experimental design for anticonvulsant activity

The experimental mice for anticonvulsant activity test were divided into 20 groups each consisting of 7–10 mice of either sex (equal ratio). All synthesized coumarin derivatives were suspended in 0.5% methylcellulose and given using oral gavage for once (10 ml/kg). Group A is model control group was given vehicle; Group B is positive control group was given carba-

mazepine (100 mg/kg (Bhat and Al-Omar, 2011; Alrohaimi et al., 2014) suspended in 0.5% methylcellulose. All coumarin derivatives were given to mice in three different dosages (30, 100, and 300 mg/kg (Alrohaimi et al., 2014) for evaluating their anticonvulsant activities. Treatment groups were entitled with the same group name as in the anti-inflammatory activity test such as Group **8a1-8a3** are given treatment 1 in low, middle, and high doses, respectively.

2.3. Induction of inflammation and assessment of anti-inflammatory activity

The acute anti-inflammatory activities of six different coumarin derivatives were investigated and compared in rats with carrageenan-induced paw edema method (Taşkın et al., 2019). Briefly, rats were fasted for 12 h with free access to water until the experiment starts and then, different groups were treated with 6 new coumarin derivatives at 3 different dosages (5, 10, 25 mg/kg, p.o.), indomethacin (10 mg/kg, p.o.), and vehicle (0.5% methylcellulose; 5 ml/kg, p.o.) 30 min before the administration of carrageenan (Taskin et al., 2018). In order to induce inflammation, 0.1 ml of 1% (w/v) carrageenan in saline was injected subplantarily in the right paw of Sprague-Dawley rats (Taskin et al., 2020). The volume of the edema development and its duration was determined for 5 h using a digital plethysmometer at times 0, 1, 2, 3, 4, and 5 h after carrageenan injection (Tamrat et al., 2017). The percent inhibition of edema was measured in comparison to the control animals and was calculated according to the following formula (Taskin et al., 2020): % Inhibition = $[(V_t - V_o)_{\text{control}} - (V_t - V_o)] / [(V_t - V_o)_{\text{control}}] \times 100$. Where V_t is the paw volume at time t , V_o is the paw volume before carrageenan injection, $(V_t - V_o)$ is edema in paw after time ' t '.

2.4. Induction of epileptic seizure and assessment of anticonvulsant activity

The anticonvulsant activities of six different coumarin derivatives were investigated and compared by PTZ-induced seizures model in mice (Kamiński et al., 2020). This rodent model is widely utilized as a standard method for predicting protection against tonic-clonic seizures in humans (Keshavarz and Yekzaman, 2018). Effective dose 50 (ED_{50}) value for PTZ (80 mg/kg) was calculated by the method of Litchfield and Wilcoxon (Litchfield and Wilcoxon, 1949). Firstly, mice were fasted overnight and then, carbamazepine (100 mg/kg, p.o.), vehicle (0.5% methylcellulose; 5 ml/kg, p.o.), and all coumarin derivatives at 3 different dosages (30, 100, and 300 mg/kg (Arshad et al., 2014)) were administered to different groups 30 min prior to the administration of PTZ (80 mg/kg, i.p. (Pithadia et al., 2013)). After PTZ injection, each mice was immediately placed in a separate observation box and their behaviors were observed for 30 min for frequency of convulsions, latency to the onset of seizures, percentage of grades, and mortality (Khoshnood-Mansoorkhani et al., 2010). The animals that survived after that period of time were considered to be protected. Moreover, each seizure was graded according to a modified Racine scale as follows: 1-No movements; 2-Head nodding and myoclonic jerks (MKJ); 3-Forelimb clonus; 4-Rearing; 5-Falling and generalized convulsions with tonic extension (Racine, 1972).

2.5. Statistical analysis

All data were expressed as mean $\pm$ standard error of mean (SEM). Statistical analysis was performed using GraphPad Prism 6.0 software (GraphPad Software, Inc., San Diego, CA). Groups of data were compared with analysis of variance (ANOVA) and Dunnet *post-hoc* test. Values were considered significantly different at $p < 0.05$.

3. Results And Discussion

3.1. Chemistry

The synthesis of the diacyl hydrazide-coumarin derivatives **8 a-f** (Table 1) was performed by the reaction of 3-carbonyl chloride coumarin **7** with some substituted aryl acid hydrazides **3a-f** as indicated in Scheme 1. To synthesis the aryl acid hydrazides at first aryl acids **1a-f** were converted into methyl esters by refluxing with methanol in sulfuric acid in good to excellent yields (65–99%), and then to corresponding hydrazides **3a-f** were obtained by stirring at room conditions for 16–20 h (67–95%) via hydrazinolysis. The acid chloride precursor **7** prepared in two steps. At first 2-oxo- 2H-chromene-3-carboxylate **6** was synthesized by Knoevenagel condensation of salicylic aldehyde and diethylmalonate in basic media with 90% yield and then acylated with thionyl chloride to get the key starting molecule **7** with excellent yield (95%). Finally, the target compounds, diacyl hydrazide-coumarin derivatives **8 a-f** were synthesized in good–excellent yields (70–95%) from the reaction of acid hydrazides (**15–20**) and 2-oxo-2H-

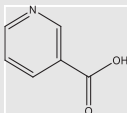
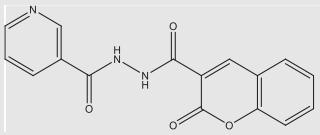
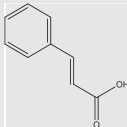
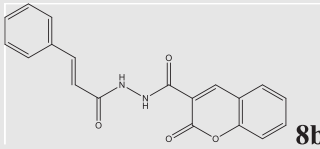
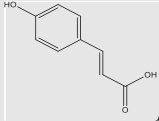
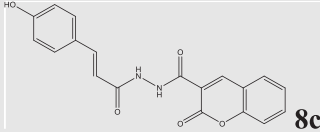
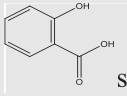
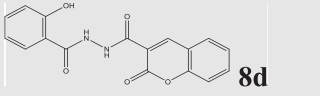
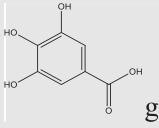
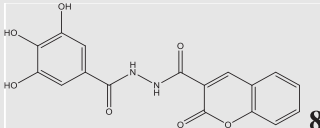
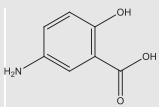
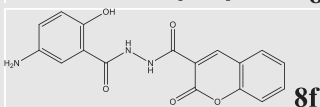
chromen-3-carbonyl chloride (**25**). All these molecules are novel except **8d** (Jesumoroti et al., 2019). However, anti-inflammatory and anticonvulsant activities of these all compounds were investigated in this study, as a first time.

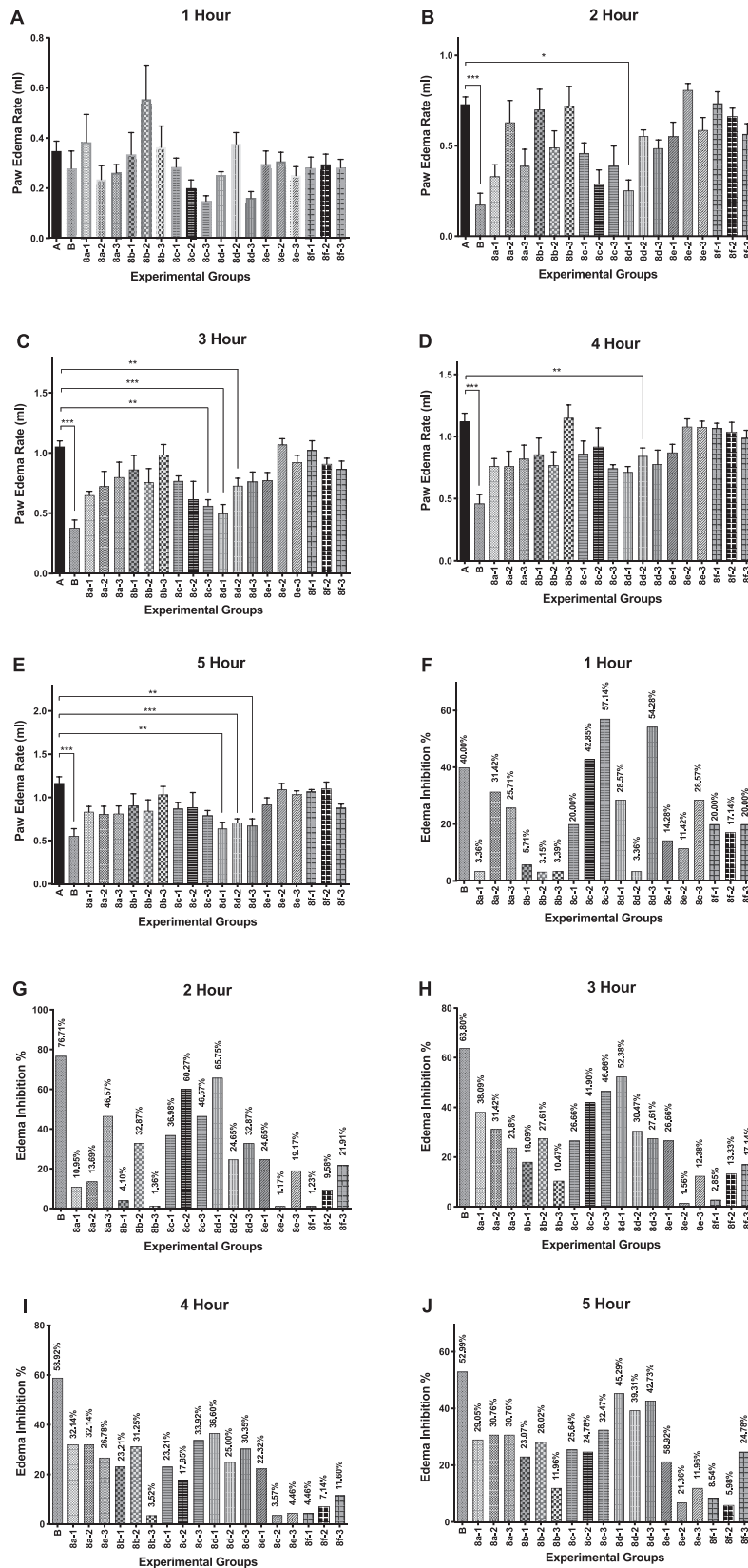
3.2. In vivo animal test

3.2.1. Anti-inflammatory activity

Inflammation is the natural defense mechanism of the body against irritants, trauma, pathogens, or microbial invasion. It occurs in the human body with symptoms such as pain, swelling, redness, heat, and disturbance of function (Aghasafari et al., 2019; Minhas et al., 2017; Pahwa and Jialal, 2019). Migration and activation of leukocytes cause tissue destruction and enhanced blood vessel permeability. Hence, inflammation could be occur by activation of several enzymes (lipoxygenases and cyclooxygenases (COX)), production of inflammatory mediators (leukotrienes and prostaglandins), and reactive oxygen species (Abdel-Lateff et al., 2020). In the present study, 0.1 ml of 1% (w/v) carrageenan that is commonly used to induce an acute model of inflammation was injected subplantarily in the right paw of rats. Carrageenan shows its paw edema effects in two phases: The initial phase (0–1 h) is related to the release of serotonin, bradykinin, and histamine; and the second phase (from the 2nd hour) is derived polymorphonuclear leucocytes infiltration and high production of prostaglandins. Amounts of free radicals, tumor necrosis factor- α , reactive oxygen species, interleukin-1 β , and nitric oxide also increase because of releasing polymorphonuclear leucocytes (Chauhan et al., 2018; Pasqua et al., 2019). Indomethacin that

Table 1 The synthesis of the diacyl hydrazide-coumarin derivatives **8 a-f**.

Entry	Aryl acid	Corresponding hybrid	Yield (%)
1	 nicotinic acid	 8a	90
2	 cinnamic acid	 8b	85
3	 <i>p</i> -Coumaric acid	 8c	70
4	 salicylic acid	 8d	90
5	 gallic acid	 8e	95
6	 5-Aminosalicylic acid.	 8f	80



is a nonsteroidal anti-inflammatory drug (NSAID) was used as being the positive control group in this study. The mechanism of effect of this drug is to reduce the production of prostaglandins and prevent the production of prostaglandins from arachidonic acid (Lucas, 2016).

In order to screen the anti-inflammatory profile of the synthesized diacylhydrazide-coumarin derivatives of aryl acids (**8a-f**) were tested for 5 h at 3 different dosages (5, 10, 25 mg/kg). The changes in the level of paw edema volumes and percentage inhibition of all groups were shown in Fig. 3. According to the results, the anti-inflammatory effects of all tested compounds were dose dependent. At 1st h, the maximum percentage inhibition of carrageenan-induced paw edema volumes was shown by the compound **8c** with the highest dose (25 mg/g dose; 57.14%) while the compound **8d** with the lowest dose (5 mg/kg) was more efficient at 2nd, 3rd, 4th, and 5th hours (65.75%; 52.38%; 36.60% and 45.29%, respectively) when compared to the control group (Fig. 2).

Indomethacin significantly caused inhibition of carrageenan-induced paw edema volume at 1st, 2nd, 3rd, 4th, and 5th hours, (40.00%, 76.71%, 63.80%, 58.92%, 52.99%, respectively) compared to the control group. The results indicate that at 1st h, the anti-inflammatory effect of **8c** with dose 25 mg/kg (57.14%) was greater than the anti-inflammatory effect of indomethacin (40.00%). However, among the tested compounds, **8d** with the lowest dose has the maximum average inhibition of paw edema volume during five hours. Furthermore, the anti-inflammatory effects of **8c**, and **8f** increased with an increase in applied dose according to percentage inhibition of paw edema volumes. When we consider about structure and activity relationship, it's clearly seen that hydrophilic substituents on aryl sub-units of diacyl-coumarin molecules significantly decreases the inhibition of carrageenan induced paw edema volume. Hence, as we expected, compounds **8e** and **8f** were non-efficient molecules for the percentage inhibition of carrageenan-induced paw edema volumes.

3.2.2. Anticonvulsant activity

Within the tests utilized for the assessment of anticonvulsant activity, the PTZ test is of predictive relevance about the clinical spectrum of activity of experimental compounds, because the PTZ test is assumed to identify anticonvulsant drugs effective against human generalized tonic-clonic seizures (Khoshnood-Mansoorkhani et al., 2010). PTZ is a noncompetitive antagonist of GABA_A receptors that acts through the *tert*-butyl-bicyclo-phosphorothionate site of the receptor and decreases its activity. Another assumption for the mechanism of PTZ is to change the potassium and calcium channel conductance (Asadi-Shekaari et al., 2014). GABA is a major inhibitory neurotransmitter in the brain and the inhibition of

GABA is the underlying factor in epilepsy (Gowda et al., 2012).

Currently used anticonvulsant drugs are effectively control epileptic seizures in almost 75% of the patients. Moreover, treatment difficulty is caused by undesirable side effects from the drugs used clinically; so new antiepileptic drugs have greater potency than more established anticonvulsant drugs. Therefore, newly synthesized diacyl hydrazine- coumarin derivatives are being seriously investigated for their antiepileptic potentials (Alrohaimi et al., 2014; Khoshnood-Mansoorkhani et al., 2010; Zhu et al., 2014). Carbamazepine, as a standard anticonvulsant drug, shows its antiepileptic effects via enhancing of sodium channel inactivation by reducing high-frequency repetitive firing of action potentials and action on synaptic transmission (Tolou-Ghamari et al., 2013). It has been widely used as standard drug in anticonvulsant activity studies and we also used carbamazepine as reference anti-epileptic drug in our study.

In this study, the anticonvulsant potential of six newly synthesized diacyl hydrazine- coumarin derivatives with three different doses was investigated by PTZ model. Compounds (*E*)-*N'*-cinnamoyl-2-oxo-2H-chromene-3-carbohydrazide (**8b**) and (*E*)-*N'*-(3-(4-hydroxyphenyl)acryloyl)-2-oxo-2H-chromene-3-carbohydrazide (**8c**) in low doses and *N'*-(2-hydroxybenzoyl)-2-oxo-2H-chromene-3-carbohydrazide (**8d**) in all doses have significant anticonvulsant potential against PTZ-induced convulsions (Fig. 3). PTZ-induced seizure model in the discovery of new antiepileptic drugs has advantages such as using of intact rodents as easy models that detect anticonvulsant effects regardless of the mechanisms of action. PTZ test can be used in high throughput screening according to the National Institutes of Health Anticonvulsant Screening Program. Besides, PTZ model can provide insight into pharmacokinetic-pharmacodynamic relations, which are so valuable for human studies (Löscher, 2011; Rho and White, 2018).

PTZ (80 mg/kg, i.p.) induced tonic-clonic seizures in all the animals used. All experimental groups were compared with the model control group. Mice pretreated with the diacyl hydrazide-coumarin derivatives at three different doses of 30, 100, and 300 mg/kg p.o. were compared for the onset of convulsion, percentage of grades, and mortality. Compounds **8b**, **8c** and **8d** with the lowest dose (30 mg/kg) and **8d** with medium dose (100 mg/kg), significantly delayed the onset of convulsion ($p < 0.05$) compared to control group (1.07 ± 0.04). The results are given in Fig. 3; **8b-1** (2.41 ± 0.26), **8c-1** (2.10 ± 0.54), **8d-1** (2.09 ± 1.03), and **8d-2** (2.16 ± 1.03). On the other side, survival was significantly increased by group B (75%); **8b-1** (86%); **8c-1** and **8c-3** (71%); **8d-1**, **8d-2**, and **8d-3** (71%, 86%, 86%, respectively), and **8f-3** (71%) compared to the control group (50%). The standard anti-epileptic drugs, carbamazepine (100 mg/kg) inhibited the severe tonic-clonic

Fig. 2 Paw edema rate (ml) and % inhibition of paw edema on carrageenan-induced inflammation model ($n = 6$). The paw edema volumes were the paw volumes' difference measured between at different time and at zero hour: (A) 1h, (B) 2h, (C) 3h, (D) 4h, and (E) 5h. The edema inhibitions % were analyzed at determined different times by comparing to the control group: (F) 1h, (G) 2h, (H) 3h, (I) 4h, and (J) 5h. Values were expressed in Mean $\pm$ SEM ($n = 6$ animals for each group). Two-way ANOVA was carried out followed by post hoc Dunnet multiple comparison test. Compounds were compared to the control group and statistical significance is expressed as * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

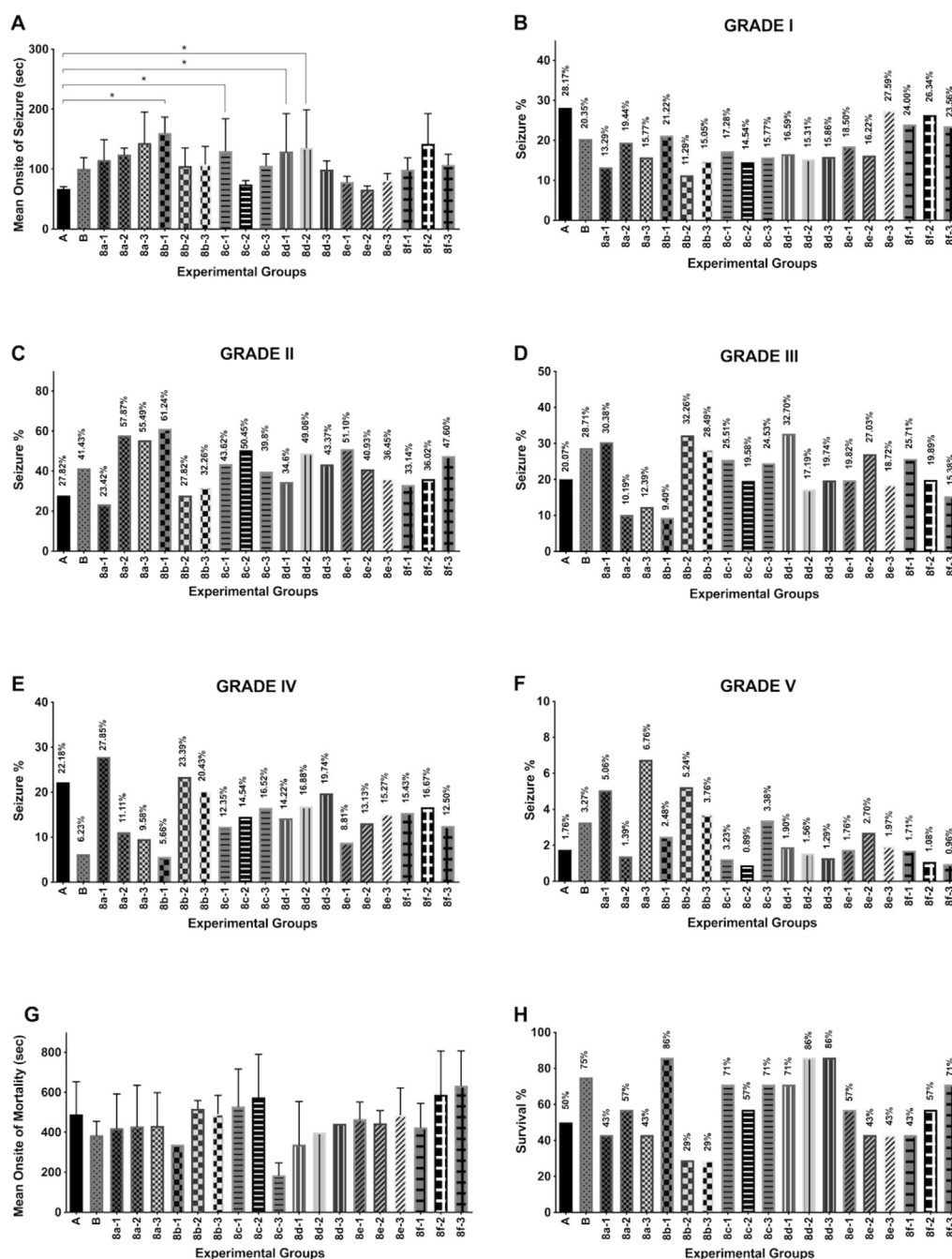


Fig. 3 Effects of coumarin derivatives on pentylenetetrazole (PTZ)- induced seizures in mice ($n = 7-10$). The mean onset of seizure (sec) (A); seizures % (grades 1-5) of experimental groups: (B) Grade I, (C) grade II, (D) grade III, (E) grade IV, and (F) grade V; the mean onset of mortality (sec) (G) and survival % (H) were analyzed. Values were expressed in Mean $\pm$ SEM ($n = 7-10$ animals for each group). One-way ANOVA was carried out followed by post hoc Dunnet multiple comparison test. Compounds were compared to the control group and statistical significance is expressed as $*p < 0.05$.

convulsions, especially considerably decreased the grade IV to 6.23% compared to control group (22.18%) and mortality in mice (Fig. 3) although it could not delay the onset of convulsion. It was also observed that although the survival rate of carbamazepine is 75%, half of animals in this group had long-term grade 5 activity during more than half of observation period. Compound **8b** which was donated with two hydrophobic aryl units, showed a strong anticonvulsant effect

in low dose but this effect was reversed in higher doses. The survival ratio decreased to 29% in middle and high doses. When we look at overall, the only compound **8d** has the anticonvulsant potential in a dose-dependent manner. The mean onsite of mortality increased in a dose-dependent manner. Also, compounds **8b** and **8c** may have anticonvulsant potential in low dose while compound **8f** has a potential in a dose-dependent manner according to the mean onsite of mortality

and survival ratio. In these treatment groups, the ratio of severe grades and mortality decreased while the onset of seizure and mortality increased. The anticonvulsant activity of these treatment groups may be explained by inhibited and/or attenuated PTZ-induced seizures of the mice by enhancing GABA-mediated inhibition and/or, decreasing the excitatory neurotransmission and/or, by blocking the sodium channels and/or by neutralizing the PTZ binding site. However, the exact mechanism(s) in anticonvulsant and anti-inflammatory effects will have to be elucidated in future studies. From these data, ideas for future molecular modification leading to compound with greater favorable pharmacological properties may be derived.

4. Conclusion

We have successfully synthesized and evaluated a number of novel diacylated hydrazide-coumarin derivatives as anti-inflammatory and anticonvulsant agents. Among these molecules, *N'*-(2-hydroxybenzoyl)-2-oxo-2*H*-chromene-3-carbohydrazide founded has a potential anti-inflammatory effect against carrageenan-induced paw edema model and anticonvulsant efficacy against the tonic-clonic seizure type in rodents. However, further studies are needed to bring out more clearly the facts concerning the exact mechanisms in which *N'*-(2-hydroxybenzoyl)-2-oxo-2*H*-chromene-3-carbohydrazide attenuates the carrageenan-induced paw edema and PTZ-induced seizures.

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Appendix A. Supplementary material

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.arabjc.2020.10.034>.

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